WEST

Help | Logout | Interrupt

Main Menu | Search Form | Posting Counts | Show S Numbers | Edit S Numbers | Preferences

Search Results -

Terms	Documents
(E near0 coli near0 PolyA near0 polymerase)	0

Database:
- US Patents Full-Text Database
- US Pre-Grant Publication Full-Text Database
- JPO Abstracts Database
- EPO Abstracts Database
- Derwent World Patents Index
- IBM Technical Disclosure Bulletins

Refine Search: (E near0 coli near0 PolyA near0 polymerase) Clear

Search History

Today's Date: 8/9/2001

Search (bacteria? (w) PolyA polymerase)

WEST

Help | Logout | Interrupt

Main Menu | Search Form | Posting Counts | Show S Numbers | Edit S Numbers | Preferences

Search Results -

Terms	Documents
l5 same kit	1

Database: US Patents Full-Text Database
US Pre-Grant Publication Full-Text Database
JPO Abstracts Database
EPO Abstracts Database
Derwent World Patents Index
IBM Technical Disclosure Bulletins

Refine Search: l5 same kit Clear

Search History

Today's Date: 8/9/2001

DB Name	Query	Hit Count	Set Name
USPT	l5 same kit	1	L6
USPT	l4 same polymerase	4	L5
USPT	l2 same fluorescen$	9	L4
USPT	label$ near0 RNA	1185	L3
USPT	label$ near0 ribonucleotide	58	L2
USPT	label near0 ribonucleotide	2	L1

WEST

Generate Collection

L19: Entry 1 of 2 File: USPT May 8, 2001

DOCUMENT-IDENTIFIER: US 6228638 B1
TITLE: Escherichia coli CSRB gene and RNA encoded thereby

DEPR:
cDNA was prepared by treating csrB RNA (5 .mu.g) with polyA polymerase (4 units) in a reaction containing 250 .mu.M ATP, 40 mM Tris HCl pH 8.0, 10 mM MgCl.sub.2, 2.5 mM MnCl.sub.2, 250 mM NaCl, 1 mM DTT, followed by cDNA synthesis using the Riboclone cDNA Synthesis Systems AMV RT (Promega, Madison, Wis.) according to the manufacturer's specifications. cDNA was made blunt-ended using T4 DNA polymerase and was cloned into the SmaI site of vector pUC19 [Yanisch-Perron et al. (1985) Gene 33:103-119] using E. coli strain DH5.alpha. for transformation. Approximately 200 clones were saved, 14 of which were at least partially sequenced and mapped on the E. coli genome using database searches.

WEST

End of Result Set

Generate Collection

L19: Entry 2 of 2 File: USPT Jul 1, 1997

DOCUMENT-IDENTIFIER: US 5643766 A
TITLE: Synthesis of full-length, double-stranded DNA from a single-stranded linear DNA template

BSPR:
Also within the invention is a primer which consists of a substantially purified, ss homopolymeric oligoribo-nucleotide [e.g., poly(rG), poly(rC), poly(rU) or poly(rA)] at least 5 (preferably between 5 and 30) ribonucleotides in length. This primer may be part of a kit useful for converting a ss linear DNA strand into a ds DNA duplex (or a ss RNA strand such as mRNA into first a ss linear DNA strand and then into a ds DNA duplex), which kit might also include a first enzyme (e.g., TdT or polyA polymerase) capable of adding a homopolymeric tail to the 3' end of the ss linear DNA strand; a second enzyme (e.g., E. coli DNA Polymerase I) capable of synthesizing DNA complementary to a DNA template; and, optionally, instructions for using the kit. The kit could additionally include a preparation containing a single deoxynucleotide triphosphate (e.g., a dNTP selected from dCTP, dGTP, dATP and dTTP), which can be polymerized by the first enzyme into a homopolymeric ss DNA tail complementary to the primer provided with the kit; a mixture of four dNTPs for use with the second enzyme; a third enzyme (e.g., RNase H) capable of removing RNA from an RNA.DNA duplex without significant degradation of the DNA portion of the duplex; and, optionally, a fourth enzyme (e.g., T4 polymerase) capable of digesting a ss DNA tail attached to one strand of a ds DNA molecule.

WEST

Generate Collection Print

L10: Entry 2 of 22 File: USPT Mar 12, 2002

DOCUMENT-IDENTIFIER: US 6355423 B1
TITLE: Methods and devices for measuring differential gene expression

Detailed Description Text (233):
Another class of dyes advantageously used in the present invention are fluorescence resonance energy transfer dyes (hereinafter called "FRET" dyes). FRET dye emission results when a donor fluorophore moiety, excited by a light source, transfers its excitation energy to an acceptor fluorophore moiety, which then emits a visible photon. FRET dyes can be detected by emission of the acceptor moiety, or alternatively, by quenching of the donor moiety. The energy transfer is distance-dependent, and its efficiency decreases with increasing distance (dropping off beyond approximately 10-100 angstroms). Since FRET dyes provide fluorescence emission of intensity comparable to or greater than that of other fluorescent dyes, such dyes can be advantageously used in this invention for any fluorescent labeling moiety. See, e.g., Ju et al., 1996, Nuc. Acids Res. 24:1144-1148. Further, since FRET dyes are advantageous for investigating distance dependent effects in nucleic acids, the donor and acceptor moieties can be placed on different oligomers of this invention to indicate that a fluorescence signal comes from correctly configured nucleic acids. See, e.g., Mergny et al., 1996, Nuc. Acids Res. 22:920-928. For example, a donor can be placed on a probe and an acceptor on a stacking oligomer in order to indicate that a fluorescence signal originates only from correctly and specifically hybridized stacking oligomers. Any pair of FRET dyes suitable to conjugation of oligomers can be adapted to this invention. Many non-FRET dyes can be used in a FRET mode. For example, FAM can act as a donor for such acceptor dyes as FAM, JOE, TAMRA, and ROX. Exemplary FRET dyes are listed in Table 13 in Section 6.7.

WEST

Generate Collection

L11: Entry 3 of 7 File: USPT Nov 21, 2000

DOCUMENT-IDENTIFIER: US 6150170 A
TITLE: Method for introducing and expressing genes in animal cells, and live invasive bacterial vectors for use in the same

BSPR:
RNA polymerases are DNA-binding proteins that recognize specific sequences in the DNA of promoter regions. RNA polymerases catalyze the synthesis of RNA molecules by polymerizing nucleoside triphosphates in the specific order that is dictated by the DNA coding sequence (Libby et al, Mol. Micro., 5:999-1004 (1991); Kerppola et al, FASEB J., 5:2833-2842 (1991); Alberts et al, Molecular Biology of the Cell, Eds., Garland Publishing Inc, New York, N.Y. (1994); Watson et al, Molecular Biology of the Gene, Vol. 1, Eds., The Benjamin/Cummings Publishing Comp. Inc., Menlo Park Calif. (1987); and Lewin, supra). RNA polymerases of prokaryotes typically are composed of two identical .alpha. subunits and two similar, but non-identical, .beta. and .beta.' subunits (Ishihama, Mol. Micro., 6:3283-3288 (1992); Watson et al, supra; Alberts et al, supra; and Lewin, supra). The specificity of prokaryotic RNA polymerases for a given promoter region is mediated by specific a factors that recognize core sequences encoded by the DNA in the promoter regions (Libby et al, supra; and Lewin, supra).

WEST

Generate Collection

L5: Entry 2 of 4 File: USPT Jul 27, 1999

DOCUMENT-IDENTIFIER: US 5928905 A
TITLE: End-complementary polymerase reaction

BSPR:
In some embodiments of the invention, the polynucleotides product(s) generated thereby are labelled, such as with radioisotopic, biotinyl, or fluorescent label moieties, by incorporation of labelled ribonucleotide or deoxyribonucleotides or the like into nascent polynucleotide by polymerase-mediated catalysis.